Mitochondrial DNA Analysis Revealed Gondwanan Origin of The Common Evening Brown Butterfly, *Melanitis Leda* (Lepidoptera: Nymphalidae)

Mashhoor, K.^{1,4}, Lazar, K. V.², Shanas, S.³ and Ramesh, N.^{1*}

¹Department of Biotechnology, J.J. College of Arts and Science, Namanasamudram, Pudukkottai, Tamilnadu, 622404, India; ²Department of Environmental Science, Central University of Kerala, Nileswaram, Kasaragod, Kerala, Pin 671314, India; ³Division of Entomology, Rice ResearchStation, Kerala Agricultural University, Thekkekara, Moncompu, Alappuzha, Kerala, 688503, India; ⁴Department of Biotechnology, E. M. E. A. College of Arts and Science, Kondotti, Malappuram, Kerala, 673638, India.

Mashhoor, K., Lazar, K. V., Shanas, S. and Ramesh, N. (2015). Mitochondrial DNA analysis revealed Gondwanan origin of the common evening brown butterfly, *Melanitis leda* (Lepidoptera: Nymphalidae). International Journal of Agricultural Technology 11(3):649-656.

Abstract The common evening brown butterfly, *Melanitis leda* is widely distributed in Africa, South Asia, South-East Asia and Australia. Here we report the partial sequence of cytochrome oxidase subunit I (COI) of *M. leda* isolated from Kerala, India. Comparison of the COI sequence of *M. leda* revealed close similarity to *M. leda* of Thailand, Madagascar and Tanzania than that of Papua New Guinea and Australia. The results indicate the common origin of the Common Evening Brown butterfly in Gondwana which later evolved into two clads with the geographical drifting of the countries. Genetically, *M. leda* of India is closer to those from Madagascar than that of Australia, depicting the rate of genetic variation of the insect has a strong co-relation with the time of its separation during the period of continental drift and the changes in environmental factors.

Keyword: Cytochrome oxidase, Melanitis leda, phylogeny, Gondwana

Introduction

Melanitis leda, the Common Evening Brown butterfly seen flying at dusk, is widely distributed in Africa, South Asia and South-east Asia extending to parts of Australia. This leaf foliating insect is considered as a minor pest of paddy. Its larvae, the green-horned caterpillar, feed on the margin and tip of the leaves and remove the leaf tissue and leaf veins. The damage caused by the green-horned caterpillar is almost similar to that of grasshoppers and armyworms. Severe infestations of the larvae results in heavy chlorophyll lose

^{*} Corresponding author: Ramesh, N.; Email: rameshnjjcol@gmail.com

due to the removal of leaf tissue and in a substantial loss of paddy yield (Heinrichs and Barrion, 2004).

Cladistics analysis of mitochondrial gene (COI) and two nuclear gene sequences revealed four major clades in Nymphaledae family (Wahlberg *et al.*, 2003). Murray and Prowell (2005) reported the phylogeny and evolutionary history of the neoptropical Satyrine subtribe Euptychiina using the COI and elongation factor gene sequences. Gaikwad *et al.* (2012) studied the utility of DNA barcoding for the identification of nymphalids butterflies isolated from India. Here we report the partial sequence of COI gene of *M. leda* isolated from Kerala, India and review its possible origin and divergence.

Material and methods

The adults of Common Evening Brown butterfly, *M. leda* were collected from the paddy fields of Kerala. The genomic DNA from the larvae was isolated using GeNei Ultrapure Mammalian Genomic DNA Prep Kit (Bangalore GeNei, Bangalore) as per the Manufacturer's instruction.

The COI gene was amplified using the forward primer with DNA sequence 5' CATTGGAGATGACCAAATTTATAATG -3' and reverse primer with DNA sequence 5'- TGAAATTAATCCAAATCCAGGTAAA-3'. The PCR reaction mixture consisted of 2 nanogram of genomic DNA (1 μ l), 1 μ l each forward and reverse primers at a concentration of 10 μ M, 2.5 μ l of dNTPs (2 mM), 2.5 μ l 10X reaction buffer, 0.20 μ lTaq polymerase (5 U/ μ l) and 16.8 μ l H2O. The PCR profile consisted of an initial denaturation step of 5 min at 950C, followed by 30 cycles of 10 sec at 950C, 10 sec at 550C and 1 min at 720C and ending with a final phase at 720C for 3 min. The PCR product was column purified using UltraClean PCR Clean-up Kit (Mo Bio Laboratories, Inc. California) as per the Manufacturer's instruction and sequenced from both ends using the Sanger's sequencing method at SciGenom Laboratories Ltd., Cochin.

The forward and reverse sequences were assembled by using ClustalW (http://www.ebi.ac.uk/Tools/msa/clustalw2) after removing the forward and reverse primer sequences and the consensus was taken for the analysis. The phylogenetic genetic level identification of *M. leda* was done using BOLD Animal Identification System (http://www.barcodinglife.com) using sequence similarity and tree based identification (BOLD TaxonID Tree). The percentage of each nucleotide in the COI codon of *M. leda* was determined by MEGA5 software.

Results

The PCR amplification of the COI sequence of *M. leda* isolated from Kerala, India yielded a single product of 563bp (GeneBank Accession No. KC433403). The nucleotide composition analysis revealed that the percentage of nucleotide present in the COI sequence of *M. leda* isolated from Kerala is identical with that of *M. leda* isolated from the different locations of India (GeneBank Accession No. GU012619, GU012624, GU012554) and Thailand (GeneBank Accession No. HQ962136, HQ962145, HQ962140). It showed +2% variation in the composition of nucleotide 'C' and -2% variation in codons third position compared to *M. phedima* (HQ962137, HQ962138).

The nucleotide sequence analysis of *M. leda* revealed the presence of conserved nucleotide sequences in specific locations. The nucleotide 'C' in the 14th, 38th and 218th positions, nucleotide 'G' in the 149th and 162th and nucleotide 'T' in 251th positions differentiate *M. leda* from most of the Lepidopteran species. The genetic similarity study showed that there is 97.19-100% similarity with the *M. leda* isolated from India and from the geographically isolated locations like Madagascar, Tanzania, Thailand, Kenya, Gabon Papua New Guinea and Australia. The COI sequence of *M. leda* isolated from Kerala showed 100-99.80% similarity with species of *M. leda* isolated from the other parts of India, Thailand, Madagascar and Tanzania. The *M. leda* isolated from Kerala has 94.57%, 94.38%, 92.13% and 91.76% similarity with *M. costantia, M. phedima, M. amabilis*and *M. zitenius* respectively (Fig. 1).

The phylogeny tree constructed (Fig. 2) by BOLD Animal Identification System (http://www.barcodinglife.com) clearly depicts the phylogenetic position of *M. leda* isolated from Kerala. The *M. leda* isolated from different parts of the world are arranged in 2 clads. The *M. leda* isolated from India, Madagascar, Tanzania, Thailand, Kenya, China and Gabon form one clad and the *M. leda* isolated from the Papua New Guinea and Australia form another clad.

Discussion

The genetic structure analysis of *M. leda* clearly reveals its phylogenetic status and origin. The variation in the COI sequence within the same species will be very low averaging 0.43% while congeneric species possess higher sequence divergence averaging 7.7% (Hebert *et al.*, 2009). *M. leda* isolated from Papua New Guinea and Australia showed higher variation (2.43-2.66%).

This high intraspecific variation is may be due to the geographical isolation of this population.

		Dee	bability of	100	-							
Taxonomic Level	Taxon Assignme		ement (%)	98								
Phylum	Arthropoda		100	%) × 96								
Class	Insecta		100	Similarity (%) 66 96								
Order	Lepidoptera		100	92 90								
Family	Nymphalidae		100					4 45 56				89 100
Genus	Melanitis		100		88	23	34		56	67		
99 Matches :												ed Matches ay option: To
Phylum	Class	Order	Family	G	nus		Spec	ies		Simila	arity (%)	Stati
Arthropoda	Insecta L	epidoptera	Nymphalidae	Melanitis			leda			1	100	Publish
Arthropoda	Insecta L	epidoptera	Nymphalidae	Me	Melanitis		leda			,	100	Publish
Arthropoda	Insecta L	epidoptera	Nymphalidae	Me	Melanitis lede		a		1	100	Publish	
Arthropoda	Insecta L	.epidoptera	Nymphalidae	Me	Melanitis leda		a		,	100	Early-Re	
Arthropoda	Insecta L	.epidoptera	Nymphalidae	Melanitis			leda			1	100	Early-Re
		.epidoptera	Nymphalidae		anitis		ledahelena				100	Early-Re
Arthropoda		epidoptera	Nymphalidae	Melanitis			ledahelena				100	Early-Re
Arthropoda		epidoptera			Melanitis			ledaleda			100	Publish
		.epidoptera			Melanitis		ledaleda				100	Publish
		.epidoptera			Melanitis		ledaleda				100 9.81	Publish
		epidoptera. epidoptera			Melanitis Melanitis		leda leda				9.81	Early-Re Early-Re
		epidoptera.			Melanitis		ledahelena				9.81	Early-Re
Arthropoda		epidoptera			Melanitis		ledahelena				9.81	Early-Re
		epidoptera	Nymphalidae		anitis		ledaleda			99.81	Publish	
		epidoptera	Nymphalidae	Me		leda leda			99.81	Early-Re		
		epidoptera	Nymphalidae		Melanitis					9.81	Early-Re	
Arthropoda	Insecta L	epidoptera	Nymphalidae		Melanitis		leda			9	9.81	Early-Re
Arthropoda	Insecta L	.epidoptera	tera Nymphalidae		Melanitis		leda			9	9.81	Early-Re
Arthropoda	Insecta L	.epidoptera	Nymphalidae		Melanitis		leda			9	9.81	Early-Re
Arthropoda	Insecta L	.epidoptera	Nymphalidae		anitis		ledahe				99.8	Early-Re
		.epidoptera	Nymphalidae		anitis		led				7.57	Early-Re
		epidoptera	Nymphalidae		anitis		led				7.57	Early-Re
		.epidoptera	Nymphalidae		anitis		led				7.57	Early-Re
		.epidoptera	Nymphalidae		anitis		led				7.57	Early-Re
Arthropoda Arthropoda		.epidoptera .epidoptera	Nymphalidae Nymphalidae		anitis anitis		leda leda				7.57 7.57	Early-Re Early-Re
		epidoptera.	Nymphalidae		anitis		leda				7.57	Early-Re
		epidoptera	Nymphalidae		anitis		leda				7.57	Early-Re
		epidoptera	Nymphalidae		anitis		leda				7.57	Early-Re
		.epidoptera	Nymphalidae		anitis		led				7.57	Early-Re
Arthropoda		epidoptera	Nymphalidae	Me	anitis		led	a		9	7.57	Early-Re
Arthropoda		epidoptera	Nymphalidae	Me	anitis		led	a		9	7.46	Early-Re
Arthropoda	Insecta L	.epidoptera	Nymphalidae	Me	anitis		led	a		9	7.38	Early-Re
Arthropoda	Insecta L	.epidoptera	Nymphalidae		anitis		led				7.38	Early-Re
Arthropoda	Insecta L	.epidoptera	Nymphalidae		anitis		led				7.34	Early-Re
		epidoptera	Nymphalidae		anitis		lede				7.27	Publishe
		epidoptera	Nymphalidae		anitis		leda				7.19	Publishe
		epidoptera	Nymphalidae		anitis		constantia hedimaabdullae			4.57	Early-Re	
		epidoptera	Nymphalidae		anitis						4.38	Publishe
		epidoptera	Nymphalidae				phedimaabdullae			4.38	Publishe	
		epidoptera opidoptera	Nymphalidae Nymphalidae		Melanitis Melanitis		phedimaabdullae phedima				4.38 4.38	Publishe Publishe
		epidoptera epidoptera	Nymphalidae								4.38 4.38	Early-Re
		epidoptera	Nymphalidae		Melanitis Melanitis		phedima phedima				4.38	Early-Re
		epidoptera	Nymphalidae		Melanitis		phedima phedima				4.38	Early-Re
Construction of the second second		epidoptera	Nymphalidae		Melanitis		pheaima phedima				4.38	Early-Re
Arthropoda				In CI			pircul					-any ite

Figure 1. Percentage of genetic similarity of *M. leda* isolated from Kerala, India with insects isolated from other locations (generated by BOLD Animal Identification System, http://www.barcodinglife.com)



Figure 2. A portion of BOLD TaxonID Tree showing the phylogenetic relationship of *M. leda* isolated from Kerala.

Before the evolution of the Asia, Africa and Australia, India was the part of supercontinent Gondwana, which includes Australia, Antartica, South America, Newzealend, New Caledonia, New Guinea, Arabia, Iran, Africa and

Madagascar. The break-up of the supercontinent started in ~170 Million years ago (Ma), following the drifting of South America–Africa from Madagascar, India, Antarctica and Australian plate. About 132 Ma Australia-Antarctica began to drift away from India–Madagascar (Powell et al., 1988; Muller et al., 2000; Brown et al., 2003) and 90-85 Ma Madagascar separated from Indian plate (Storey et al., 1995; Torsvik et al., 2000). New Guinea separated from Australia about 30Ma. The sequential breakup of the supercontinent Gondwana in different geographical time caused the successive division of an ancestral biota (Sanmartin&Ronquist, 2004). M. leda is distributed in Asia and Africa, Australia and Papua New Guinea (Dale, 1994, Heinrichs&Barrion, 2004). Interestingly most of the geographical areas of *M. leda* distribution were the part of Gondwana except some parts of Asia. The similarity between the fauna of India and Africa prove their common origin from Gondwanaland (Krause & Maas, 1990; Zardoya et al., 1996; Cracraft, 2001; Cooper et al., 2001; Biju&Bossuyt, 2003; Noonan &Chippindale, 2006). The genetic similarity analysis and geographical distribution of *M. leda* also indicate its origin from the supercontinent Gondwana. After the separation, India continued its rapid northward migration and colliding with southern Asia (Barron & Harrison, 1980; Lee & Lawver, 1995). Conti et al. (2002) argued that Crypteroniaceae are from Gondwanan origin and they reached in the tropical Asia by transportation from Gondwana on the Indian "raft". Likewise the M. leda seen in China and Thailand may have come from the Indian subcontinent.

The Australian vegetation and climate substantially changed after its separation from Antarctica (Hill, 2004). The *M. leda* isolated from India showed more variation with the *M. leda* isolated from Papua New Guinea and Australia than Madagascar. This may be due to the early separation of Australian and New Guinea plates from the Indian plates than Madagascar plate and may be due to the changes in environmental factors of the Australian plate after its separation from the Antarctic plate. The drift between Madagascar plate and Indian plate occurred as a later event and therefore the *M. leda* isolated from the India and Madagascar showed close similarity. The results indicate that the Common Evening Brown, *M. leda* might have originated as early as the origin of the supercontinent Gondwana which later spread to the adjacent countries after the Gondwana separation.

References

Barron, E. J. and Harrison, C. G. A. (1980). An analysis of past plate motions; the South Atlantic and Indian oceans. Mechanisms of continental drift and plate tectonics. London: Academic Press. pp. 89-109.

- Biju, S. D. and Bossuyt, F. (2003). New frog family from India reveals an ancient biogeographical link with the Seychelles. Nature 435:711-714.
- Brown, B., Muller, R. D., Struckmeyer, H. I. M., Gaina, C., Stagg, H. and Symonds, P. (2003). Formation and evolution of Australian passive margins: implications for locating the boundary between continental and oceanic crust. Evolution and Dynamics of the Australian Plate. pp. 223-243.
- Conti, E., Eriksson, T., Schonenberger, K. J., Sytsma, K. J. and Baum, D. A. (2002). Early Tertiary out-of- India dispersal of Crypteroniaceae: evidence from phylogeny and molecular dating. Evolution 56:1931-1942.
- Cooper, A., Lalueza-Fox, C., Anderson, S., Rambaut, A., Austin, J. and Ward, R. (2001). Complete mitochondrial genome sequences of two extinct moas clarify ratite evolution. Nature 409:704-706.
- Cracraft, J. (2001). Avian evolution, Gondwanan biogeography and the Cretaceous-Tertiary mass extinction event. Proceedings of The Royal Society of London Series. Biological Science 268:459-469.
- Dale, D. (1994). Insect pests of the rice plant their biology and ecology. Biology and management of rice insects. New Delhi: Wiley Eastern. pp. 363-485.
- Gaikwad, S. S., Ghate, H. V., Ghaskadbi, S. S., Patole, M. S. and Shouche, Y. S. (2012). DNA barcoding of nymphalid butterflies (Nymphaledae: Lepidoptera) from Western Ghats of India. Molecular Biology Reports 39:2375-2383.
- Hebert, P. D. N., de Waard, J. and Landry, J. F. (2009). DNA barcodes for 1/1000 of the animal kingdom. Biology Letters 6:359-362.
- Heinrichs, E. A. and Barrion, A. T. (2004). Rice-feeding insects and selected natural enemies in West Africa: biology, ecology, identification. Los Banos (Philippines): International Rice Research Institute and Abidjan (Côte d'Ivoire): WARDA–The Africa Rice Center.
- Krause, D. W. and Maas, M. C. (1990). Thebiogeographical origins of the late Paleocene-early Eocene mammalian immigrants to the western interior of North America. Dawn of the age of mammals in the northern part of the Rocky Mountain interior, North America. New York: Geological Society of America. pp. 71-105.
- Lee, T. Y. and Lawver, L. A. (1995). Cenozoic plate reconstruction of the southeast Asia region. Tectonophysics 251:85-138.
- Muller, R. D., Gaina, C. and Clarke, S. (2000). Seafloor spreading around Australia Billionyear Earth History of Australia and Neighbours in Gondwanaland. Sydney: Gemoc Press. pp. 18-28.
- Murray, D. and Prowell, D. P. (2005). Molecular phylogenetics and evolutionary history of the neotropical Satyrine Subtribe Euptychiina (Nymphaledae: Satyrinae), Molecular Phylogenetics and Evolution 34:67-80.
- Noonan, B. P. and Chippindale, P. T. (2006). Dispersal and vicariance: the complex evolutionary history of boid snakes. Molecular Phylogenetics and Evolution 40:347-358.
- Powell, C. M., Roots, S. R. and Veevers, J. J. (1988). Pre-breakup continental extension in eastern Gondwanaland and the early opening of the eastern Indian Ocean.Tectonophysics 155:261-283.
- Sanmartin, I. and Ronquist, F. (2004). Southern Hemisphere biogeography inferred by eventbased models: Plant versus animal patterns. Systematic Biology 53:216-243.
- Storey, M., Mahoney, J. J., Saunders, A. D., Duncan, R. A., Kelley, S. O. and Coffin, M. F. (1995). Timing of hot-spot related volcanism and the breakup of Madagascar and India. Science 267:852-855.

- Torsvik, T. H., Tucker, R. D., Ashwal, L. D., Carter, L. M., Jamtveit, B., Vidyadharan, K. T. and Venkataramana, P. (2000). Late Cretaceous India–Madagascar fit and timing of break-up related magmatism. Terra Nova 12:220-224.
- Wahlberg, N., Weingartner, E. and Nylin, S. (2003). Towards a better understanding of the higher systematics of Nymphaledae (Lepidoptera: Papilionoidea), Molecular Phylogenetics and Evolution 28:473-484.
- Zardoya, R., Vollmer, D. M., Craddock, C., Streelmans, J. T., Karl, S. A. and Meyer, A. (1996). Evolutionary conservation of microsatellite flanking regions and their use in resolving the phylogeny of cichlid fishes (Pisces: Perciformes). Proceedings of the Royal Society of London Series B. Biological Science 263:1589-1598.

(Received: 6 January 2015, accepted: 27 February 2015)